



Monte Rosa Therapeutics

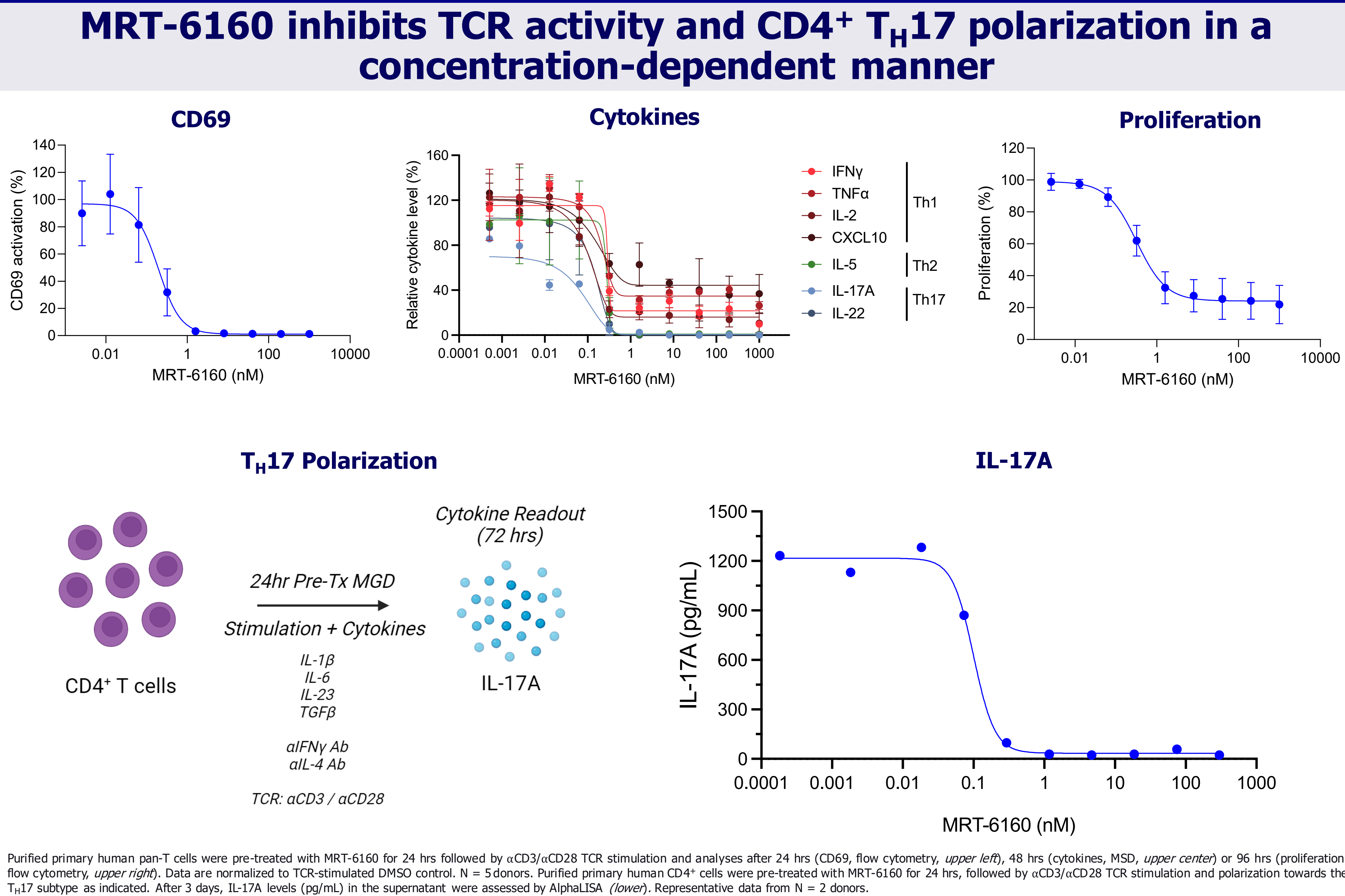
#Tu1727: MRT-6160, a VAV1-Directed Molecular Glue Degradator, Inhibits Disease Progression in a T-cell Transfer Mediated Colitis Model Concomitant with Reduced Calprotectin Expression

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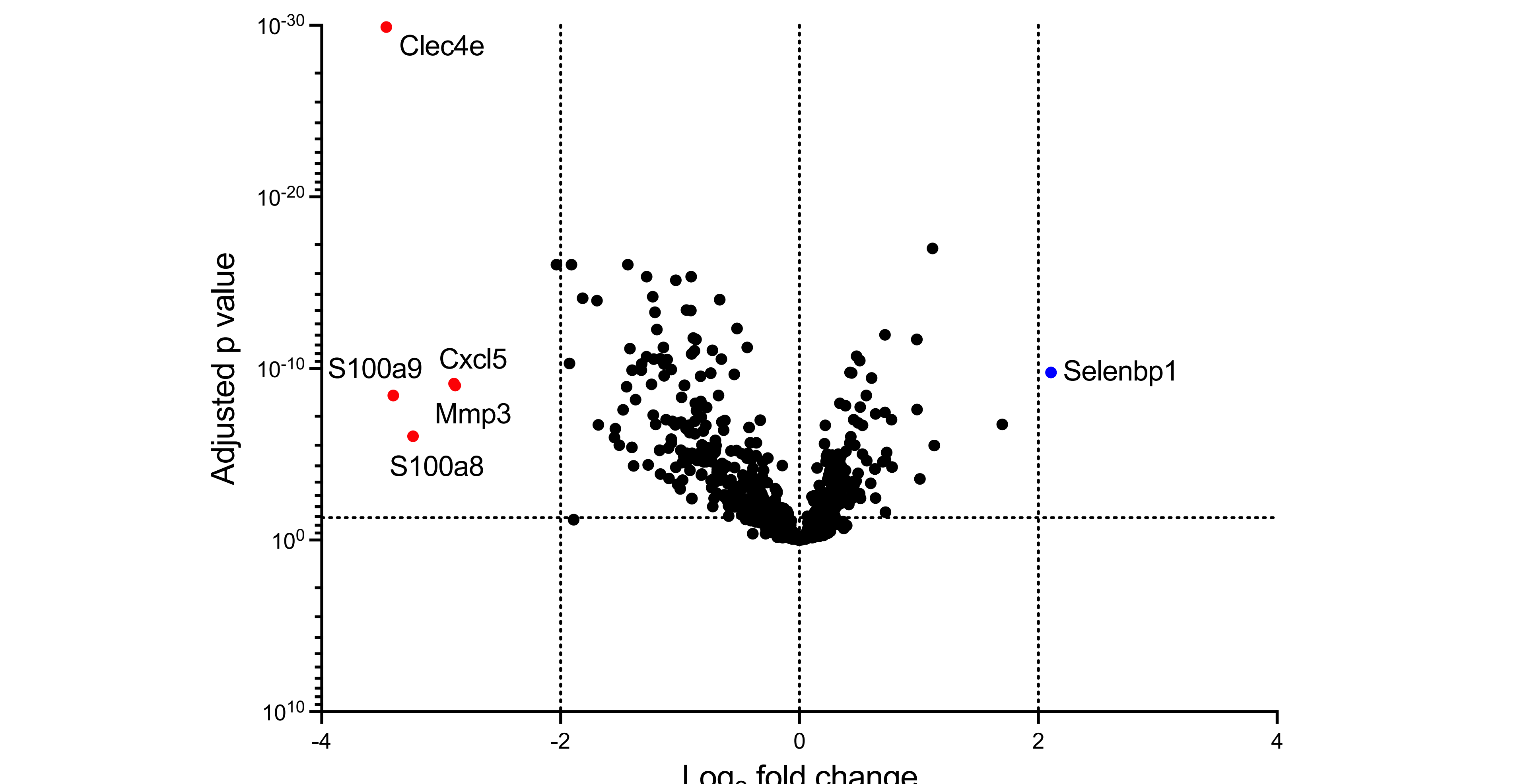
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VAV1 is a critical guanine nucleotide exchange factor in T-cell receptor signaling and activity

- VAV1 expression is highly restricted to immune cells
- VAV1 is a pivotal scaffolding protein and signaling molecule downstream of the TCR
- CRISPR-mediated KO¹ or germline genetic loss² of VAV1 is associated with decreased functions of T cells
- VAV1 degradation is predicted to impact T cell and B cell function and treat a broad set of autoimmune and inflammatory diseases

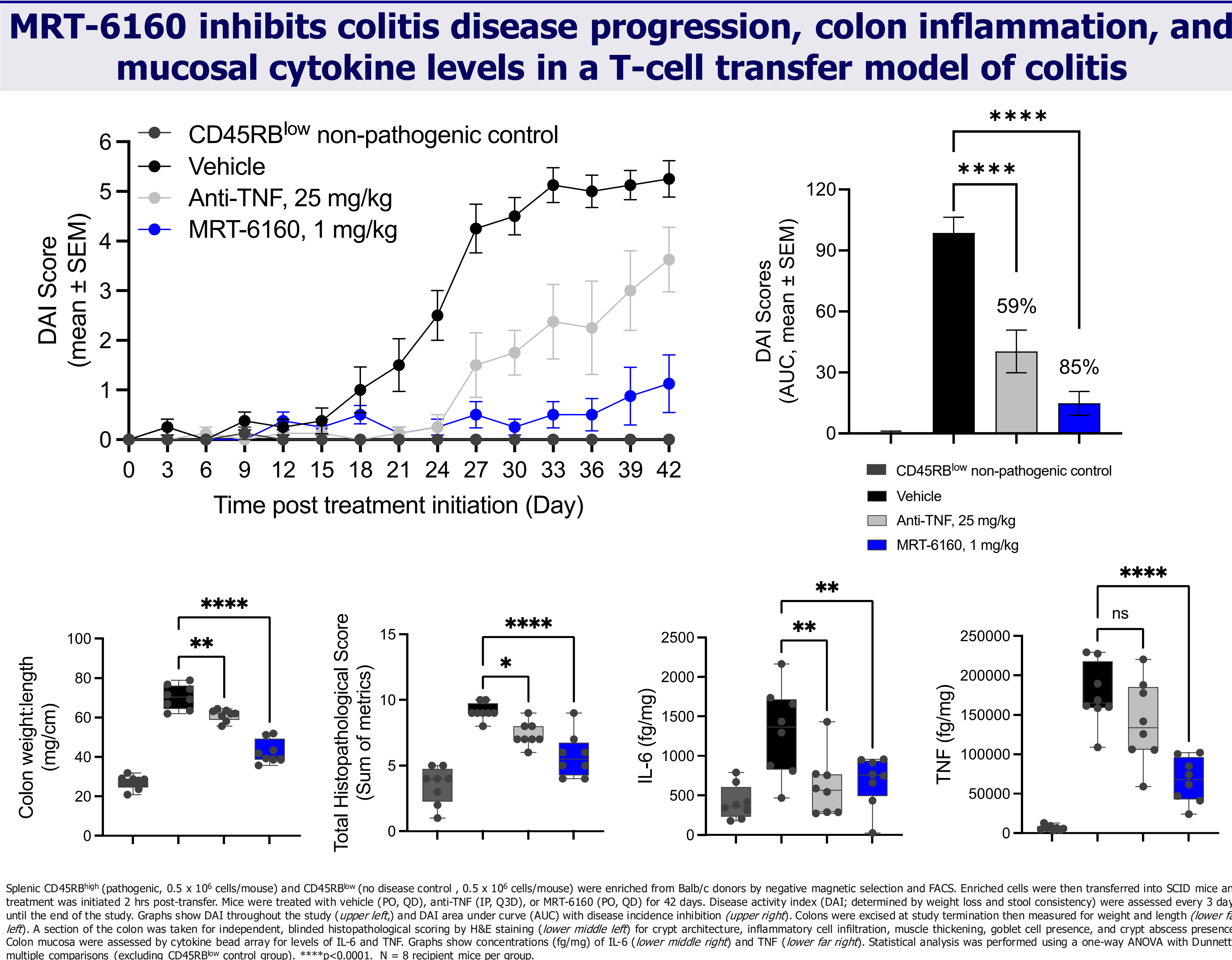


Purified primary human pan-T cells were pre-treated with MRT-6160 for 24 hrs followed by α CD3/ α CD28 TCR stimulation and analyses after 24 hrs (CD69, flow cytometry, upper left), 48 hrs (cytokines, MSD, upper center) or 96 hrs (proliferation, flow cytometry, upper right). Data are normalized to TCR-stimulated DMSO control. N = 5 donors. Purified primary human CD4⁺ T cells were pre-treated with MRT-6160 for 24 hrs, followed by α CD3/ α CD28 TCR stimulation and polarization towards the Th17 subtype as indicated. After 3 days, IL-17A levels (pg/mL) in the supernatant were assessed by AlphaLISA (lower). Representative data from N = 2 donors.

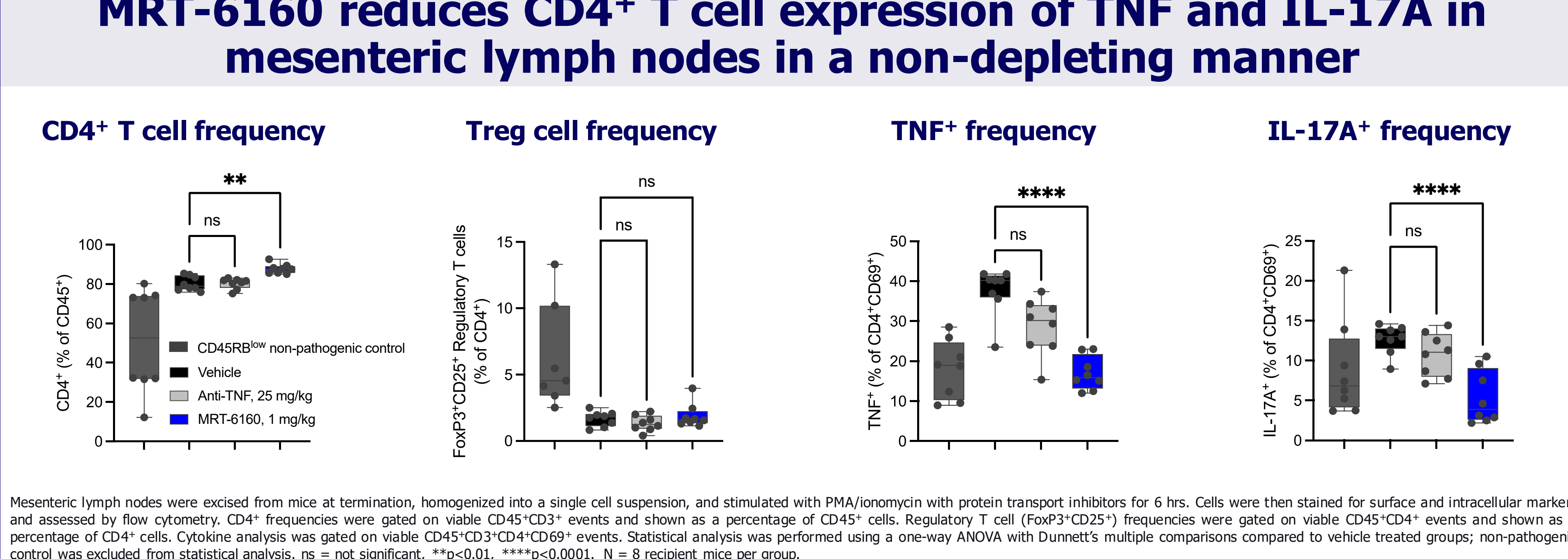


Gene	L2FC	Adj. p-value	Function
<i>Clec4e</i>	-3.46	1.2 x 10 ⁻³⁰	Induces <i>Iil1b</i> expression and T _H 17 differentiation
<i>S100a9</i>	-3.40	3.6 x 10 ⁻⁹	Calprotectin subunit, induces inflammatory activation of endothelial cells
<i>S100a8</i>	-3.24	8.8 x 10 ⁻⁷	Calprotectin subunit, induces inflammatory activation of endothelial cells
<i>Cxcl5</i>	-2.90	7.6 x 10 ⁻¹⁰	Ligand for CXCR2, induces migration and chemotaxis of T cells
<i>Mmp3</i>	-2.88	9.7 x 10 ⁻¹⁰	Cleaves extracellular matrix proteins and activates pre-cursors of TNF α and IL-1 β
<i>Selenbp1</i>	2.11	1.7 x 10 ⁻¹⁰	Selenium-binding protein, associated with long-term remission in UC patients

Colons were excised from mice at termination and stored in RNAlater. RNA was extracted from colon samples and quality was assessed by NanoDrop and Qubit. Gene counts were performed using the NanoString Autoimmune Profiling Panel and processed according to the manufacturer's instructions. Background signal was determined at twice the SD above the mean of the negative control. Data were normalized to 8 housekeeping genes and median of the ratio was used. Graphs show differentially expressed genes (DEG) in MRT-6160 vs vehicle treated mice. Table shows log₂ fold change, p-value, and description of labelled genes in volcano plot.



Splenic CD45RB^{low} (pathogenic, 0.5 x 10⁶ cells/mouse) and CD45RB^{low} (no disease control, 0.5 x 10⁶ cells/mouse) were enriched from Balb/c donors by negative magnetic selection and FACS. Enriched cells were then transferred into SCID mice and treatment was initiated 2 hrs post-transfer. Mice were treated with vehicle (PO, QD), anti-TNF (IP, Q3D), or MRT-6160 (PO, QD) for 42 days. Disease activity index (DAI), determined by weight loss and stool consistency, were assessed every 3 days until the end of the study. Graphs show DAI throughout the study (upper left) and DAI area under curve (AUC) with disease incidence inhibition (upper right). Colons were excised at study termination then measured for weight and length (lower left). A section of the colon was taken for independent, blinded histopathological scoring by H&E staining (lower middle left) for crypt architecture, inflammatory cell infiltration, muscle thickening, goblet cell presence, and crypt abscess presence. Colon mucosa were assessed by cytokine bead array for levels of IL-6 and TNF. Graphs show concentrations (fg/mg) of IL-6 (lower middle right) and TNF (lower right). Statistical analysis was performed using a one-way ANOVA with Dunnett's multiple comparisons (excluding CD45RB^{low} control group). ****p<0.0001. N = 8 recipient mice per group.



Mesenteric lymph nodes were excised from mice at termination, homogenized into a single cell suspension, and stimulated with PMA/ionomycin with protein transport inhibitors for 6 hrs. Cells were then stained for surface and intracellular markers and assessed by flow cytometry. CD4⁺ frequencies were gated on viable CD45⁺CD3⁺ events and shown as a percentage of CD45⁺ cells. Regulatory T cell (FoxP3⁺CD25⁺) frequencies were gated on viable CD45⁺CD4⁺ events and shown as a percentage of CD4⁺ cells. Cytokine analysis was gated on viable CD45⁺CD3⁺CD4⁺CD69⁺ events. Statistical analysis was performed using a one-way ANOVA with Dunnett's multiple comparisons compared to vehicle treated groups; non-pathogenic control was excluded from statistical analysis. ns = not significant, **p<0.01, ****p<0.0001. N = 8 recipient mice per group.

MRT-6160 is a rationally designed molecular glue degrader that selectively degrades VAV1

Molecular glue degraders (MGD) function to induce structural changes in ubiquitin ligases, such as cereblon, to drive the formation of ternary complexes with a non-natural target protein, termed neosubstrate.

Following binding of cereblon to the target protein, this target is then ubiquitin tagged and subsequently degraded via the proteasome-mediated degradation machinery of the cell.

MGDs can induce degradation of otherwise 'undruggable' proteins as the mechanism does not require a classical binding pocket contrary to conventional protein inhibitors, significantly increasing the target space and utility across a range of diseases.

Human PBMCs

Mouse Splenocytes

Human PBMCs and mouse splenocytes were treated for 24 hrs with 10 μ M MRT-6160 then assessed by quantitative tandem mass tag proteomics. The y-axis represents p-value (-log₁₀); the x-axis represents protein fold change (log₂) relative to DMSO (0.1%) control samples. Dark blue circles represent CXCR6 neosubstrates including the target, VAV1, and other known cereblon neosubstrates; GSPT1, IKZF1, IKZF3, CSNK1A1 (CK1 α), SALL4, and ZFP91. Purple circles represent VAV family members VAV2 and VAV3.